Aristerostoma sp.

Note: The culturing conditions below are not necessarily the optimal growth conditions for each strain, as much variation is found between strains, and cultures are not always kept in optimal growth conditions at CCAP for practical reasons. There may be more info in the individual strain data on the website.

Storing the cultures in natural daylight at room temperature should also be fine, providing they are kept out of direct sunlight.

On receipt of culture: cultures should be subcultured into fresh sterile medium as described below, ideally within a few days of receipt. If the culture vessel is very full on receipt and subculturing cannot be done immediately, we advise transferring half of the culture to a sterile container to provide air space.

ACDP Hazard Gp: 1 - Non pathogenic / non hazardous. Unlikely to cause human disease.

Culture Medium: f/2 with 1 or 2 boiled wheat grains.
Media recipes can be found on our website: www.ccap.ac.uk/index.php/media-recipes/

Lighting: mix of cool and warm white fluorescent lighting

Light Cycle: 12h light : 12h dark

Temperature: 20 degrees C

Sub Interval: 4 weeks (at CCAP, may vary depending on environment)

Culture Vessel: Tissue culture flask (30ml medium with 2 wheat grains added)

Culture Method:

Choose a dense culture from existing stocks, the state of the culture is ascertained by microscopic examination at x40 or x60. Aseptically prepare two new tissue culture flasks for each culture – add 25ml sterile medium to each new 50ml tissue culture flask.

The ciliates settle to the bottom of the flask where they feed on the bacteria; before subculturing use a sterile pipette to gently mix the culture then add around 5ml to each new flask. Add fresh boiled and cooled wheat/barley grains to encourage bacterial growth. *Aristerostoma* is also algivorous and can be fed with a few drops of *Nannochloropsis* or *Isochrysis* algae. Seal with parafilm and incubate at 20 degrees.

Use strict aseptic techniques throughout and if possible carry out all subculturing within a laminar flow cabinet (particularly important for axenic strains).