

Nss (low)

Stocks

	per 1 Litre
(1) Extra Salts	
NaNO ₃	30.00 g
Na ₂ HPO ₄	1.20 g
K ₂ HPO ₄	1.00 g
(2) Vitamin Solution	
Biotin	0.0002 g
Calcium pantothenate	0.02 g
Cyanocobalamin	0.004 g
Folic acid	0.0004 g
Inositol	1.0 g
Nicotinic acid	0.02 g
Thiamine HCl	0.1 g
Thymine	0.6 g
(3) Soil extract 1 (SE1-see overleaf)	

Medium

	per 1 Litre
Tricine	0.50 g
Stock solution 1	3.75 ml
Stock solution 2	5.00 ml
Stock solution 3 (SE1)	12.50 ml

The stock solutions are those for ASW. Make up to 1 litre with filtered natural seawater *. Adjust pH to **7.6 - 7.8** with 1M NaOH or 1M HCl prior to autoclaving. Autoclave at 15 psi for 15 minutes.

* Alternatively, use distilled water to 1 litre and 33.6 g "Ultramarine Synthetica" sea salts **.

Supply

** Waterlife Research Industries Ltd., 476 Bath Road, Longford, West Drayton, Middlesex, England, UB7 0ED. Tel: (01753) 685696.

Reviewed: 10TH August 2020

Created on: 05 Nov 2019	CCAP (Culture Collection of Algae and Protozoa), SAMS Ltd, Scottish Marine Institute, Oban, Argyll, PA37 1QA, UK Tel: +44 (0)1631 559000 Fax: +44 (0)1631 559001 Email: ccap@sams.ac.uk Web: www.ccap.ac.uk	2 Pages
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SE1 (Soil Extract 1)

Used in media for marine algae

Preparing the soil

Site selection for a good soil is very important and for most purposes a soil from undisturbed deciduous woodland is best. Sites to avoid are those showing obvious signs of man's activity and particular care should be taken to avoid areas where fertilizers, crop sprays or other toxic chemicals may have been used.

A rich loam with good crumb structure should be sought. Stones, roots and larger invertebrates should be removed during an initial sieving through a 1 cm mesh. The sieved soil should be spread to air dry and handpicked for smaller invertebrates and roots. It should be turned periodically and picked over again. When dry sieve through a finer mesh (2-4 mm) and store in an airtight container away from light and heat.

Medium

Soil is prepared as above. Air-dried soil and twice its volume of supernatant distilled water are autoclaved together at 15 psi for 2 hours and left to cool. The supernatant is then decanted and then distributed to containers in volumes suitable for making up batches of media. The aliquots and their containers are autoclaved for an appropriate length of time (e.g. 1 litre or less for 15 minutes) and are then kept in a cool place (e.g. a refrigerator) until required.

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